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Phytochemical Analysis of *Scinaia Bengalica* by GCMS

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Abstract: Marine red algae consist of various medicinal activities. Marine sources are more active than the other natural sources. One of the most important red algae is *Scinaia Bengalica*(SB) known for its phytochemical analysis by GC-MS revealed 19 chemical constituents. SB consist major constituents like oleic acid, octanoic acid, 2 hexyl-1-octanol, hexadecanol, calcitriol, bromine compounds.

Key Words: *Scinaia Bengalica*, GC-MS, calcitriol, Marine sources, phytochemical. bromine, hexadecanol.

Introduction:

Approximately 6000 species of algae present marine, most red algae are marine origin; only a few occur in freshwater. Rhodophytes are usually multicellular and grow attached to rocks or other algae, but there are some unicellular or colonial forms. Marine algae are the rich sources of structurally active and biologically active compounds (Ely et.al2004). Marine red algae consist of rich antioxidants, anticancer, antimicrobial, anti-inflammatory and antidiabetic activity (Gamal, guven et al 2010). Microalgae protein and lipoprotein found to have antibacterial, antifungal, antiviral activity (Burja AM, et al., 2001)

Sea weeds are consists of reactive antioxidant compounds such as ascorbate and glutathione(GSH), secondary metabolites including carotenoids (alpha and beta carotenoids), aminoacids, phlorotannins (phloroglucinol), tocopherols(alpha and beta tocopherols) Yuan et. al 2005). Marine algae and weeds consist of large amount of minerals and trace elements taken from the marine water and convert them in useful organic forms as they grow in a mineral rich medium.(Chapman et.al1980), They selectively absorb sodium, Potassium, Magnesium, Iodine and Bromine elements and accumulate them in its thalli. As the alkali composition sea foods prevents the blood acidosis. The marine algae consists of antibacterial and antimicrobial activity against all the tested bacteria showing maximum activity against *Bacillus subtilis*(Renughadevi et al., 2014).

Scinaia Bengalica(SB) belongs to the red algae, Rhodophyta phylum, Scinaiaacea family, Scinaia Genus, Scinaia bengalica species. Availability of SB: Madras Beach, India (Silva, Basson & Moe 1996). Detailed distribution as *Scinaia bengalica* Børgesen: Africa: South Africa (De Clerck, 2005). Red algae Rhodophyceae. South-west Asia: India (Silva, Basson & Moe 2015). *Algae of India Volume 3*. Verified by: 19 July 2011 by M.D. Guiry.

Citing AlgaeBase

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was designed to investigate phytoconstituent analysis of SB, biological property by using Gas chromatography-Mass spectrum.

Materials and Methods:

Collection and Extraction of algae:

Scinaia Bengalica was collected from the Madras Beach region, located on the Madras coastal region, Tamil Nadu, India. Cleaned marine algae were shade dried, the completely dried algae was used for further GC-MS phytoconstituent determination. The completely dried material was weighed and grind coarsely in a mechanical grinder. In the present study we evaluate the biological potencies of marine red algae.

Preparation of Ethanol Extract of SB for GC-MS Analysis:

SB was shade dried and 2g of the powdered biomass was soaked in 95% ethanol for 12hrs. Then the extract was filtered through whatman No.41 filter paper along with 0.2g of sodium sulfate to remove the sediments and traces of water in the filtrate. Before filtering, the filter paper was moistened with 95% ethanol for 12hrs. An aliquot of 2 μ l of this solution was employed for GC-MS analysis (Merlin et. al 2009).

GC-MS Analysis:

GC-MS analysis was carried out on a GC Clarus 500 Perkin Elmer system comprising of Aoc-20i auto sampler and gas chromatograph interfaced to a mass spectrometer. GC-MS instrument employed the following conditions: column-Elite-1 fused silica capillary column (30 mm x 0.25 mm ID x 1 μ Mdf), composed of 100% dimethyl poly siloxane, operating in electron impact mode at 70eV; carrier gas-helium (99.999%) at a constant flow of 1 ml/min; injection volume-0.5 μ l (split ratio of 10:1); injector temperature- 250°C and an ion source temperature of 280°C. The oven temperature was programmed from 110°C (isothermal for 2 min) with an increase of 0°C/min to 200°C, then 5°C/min to 280°C ending with a 9 min isothermal at 280°C. Mass spectra were taken at 70 eV; a scan interval of 0.5 seconds and fragments from 45 to 450 Da. Total GC running time was 36 min.

Results

GC-MS Profile of Ethanol Extract of *Scinaia Bengalica*

A high resolution mass spectrum equipped with a data system in combination with Gas Chromatography was used for the chemical analysis of active red seaweed. The crude ethanolic extract of SB based on spectral data by GC-MS analysis was found to be a mixture of volatile compounds. 19 peaks were observed with retention times as presented in Table-1.

Table 1- Phytochemicals obtained from GCMS with its retention time and peak area % of *Scinaia Bengalica*.

S.No	Retention Time	Name of the Compounds	Peak Area %
1	5.319	1,2-Dihydropyrazol-3-one, 4-(4-bromomethyl-4,	8.592
2	5.813	9, 12 octa deca dienoic acid	1.408
3	7.398	8-Pentadecanone	0.582
4	8.802	Octadecane 6-methyl	0.728
5	10.252	Hexadecanol	6.675
6	10.520	9, 12, 15-Octa deca trienoic acid	0.435
7	10.724	n-Hexa decanoic acid	2.582
8	10.997	E-11-Hexadecenoic acid, ethyl ester	2.736
9	12.135	Eicosanoic acid	28.220

10	12.956	Methyl 3-methyl-pentadecanoate	4.519
11	14.228	Z,E-2,13-Octadecadien-1-ol	0.782
12	14.500	3Trifluoroacetoxypentadecane	1.865
13	15.415	10-Bromodecanoic acid, ethyl ester	1.743
14	15.676	7-Heptadecene, 17-chloro-	1.622
15	17.814	1-Hexacosene	0.923
16	19.825	Octadecanal, 2-bromo-	4.087
17	20.565	16-Hentriacontanone	4.440
18	21.745	Calcitriol	11.339
19	23.285	Adenosine 3 Phosphoric acid	12.867

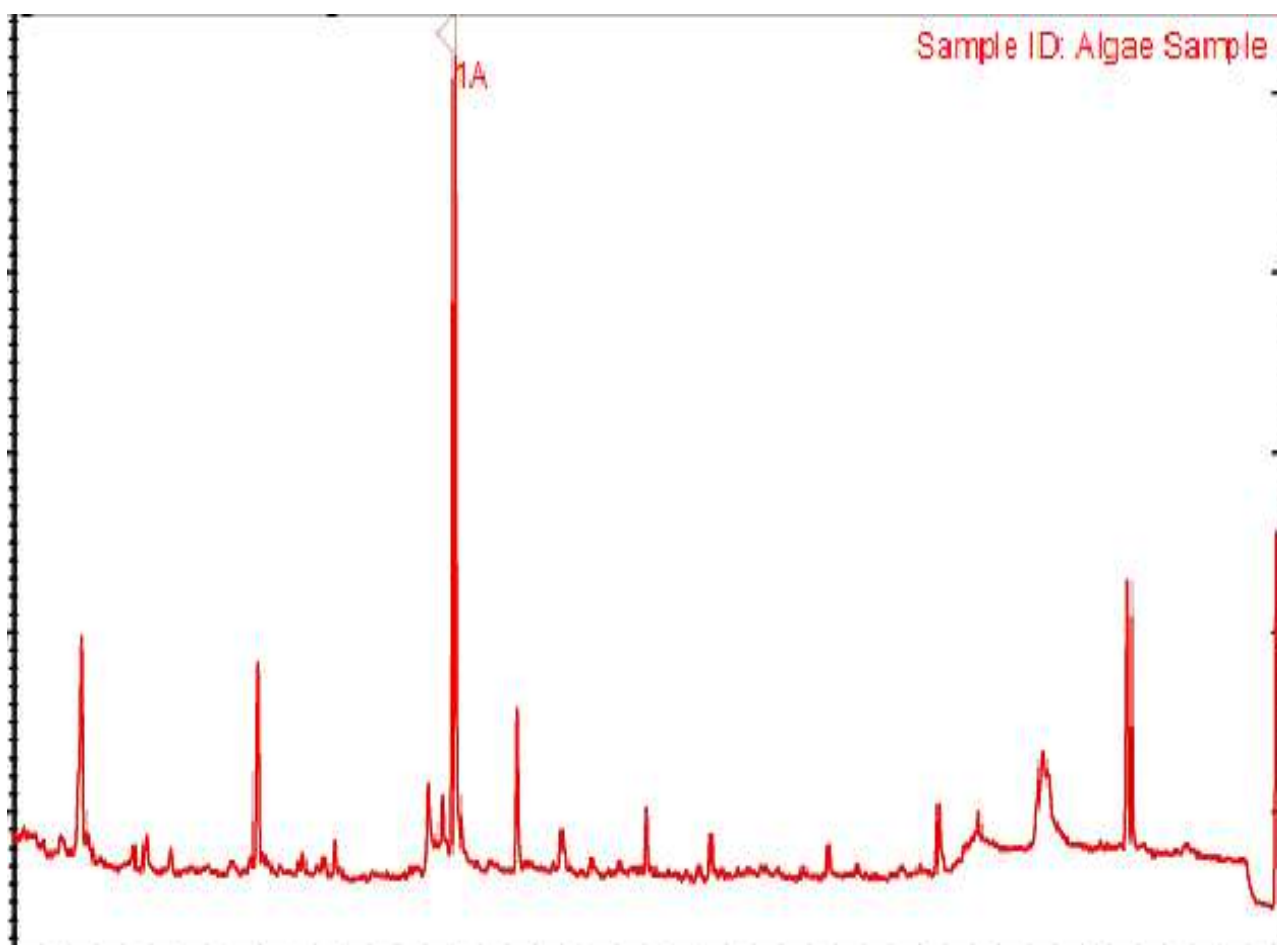


Figure 1: The phytochemical peaks obtained by Gas Chromatography Mass Spectroscopy.

The GC-MS analysis of ethanol extract of SB revealed phytoconstituent of 1,2-Dihydropyrazol-3-one, 4-(4-bromomethyl-4, 9, 12 octadecadienoic acid, 8-Pentadecanone, Octadecane 6-methyl, Hexadecanol, 9, 12, 15-Octa decatrienoic acid, n-Hexadecanoic acid, E-11-Hexadecenoic acid, ethyl ester, Eicosanoic acid, Methyl 3-methyl-pentadecanoate, Z,E-2,13-Octadecadien-1-ol, 3Trifluoroacetoxypentadecane, 10-Bromodecanoic acid, ethyl ester, 7-Heptadecene, 17-chloro, 1-Hexacosene, Octadecanal, 2-bromo, 16-Hentriacontanone, Calcitriol, Adenosine 3 Phosphoric acid.

Discussion:

Globally diabetes cases have exploded in the past two decades at 6% per annum and by the year 2025. 324 million people will be diabetic (Mohammed et.al 2007). Natural sources of drugs from marine algae are

used widely, even when their biologically active compounds are unknown, because of their effectiveness, minimal side effects in clinical experience and relatively low cost. They are one of the less explored sources of pharmacological candidates and few previous studies have found antidiabetic activities in various marine algae (Bhesh Raj et. al 2015). Marine algae inhibit the alpha-glucosidase and alpha-amylase in a uncompetitive and non-competitive manner (Murugesan et.al 2016).

Conclusion

In this study, marine microalgae have been isolated and the preliminary phytochemical analysis was done. The preliminary phytochemical analysis revealed the presence of hexadecanoic acid, eicosanoic acid, calcitriol, Bromodecanoic acid, Adenosine 3 Phosphoric acid. The ethanol extract of SB has shown more phytochemical activity than the other extracts. The marine red algae consist of various phytoconstituent that could be used as medicines after the complete research done for the identification of medicinal properties.

References

1. Ely, R., T. Supriya and C. G. Naik, 2004. Antimicrobial activity of marine organisms collected A coast of South East India. *J. Exp. Mar. Biol. Ecol.*,309: 121-127).
2. Gamal 2010, guven et al 2010, liu et al 2011, wijesekara et al 2011). PangesutiR, Kim S 2011. Biological activities and potential health benefits of sulfated polysaccharides derived from marine algae. *Carbohydrpolym* 84:14-21.
3. Burja AM, Banaigs B, Abou-Mansour E, Burgess JG and Wright PC (2001) Marine cyanobacteria - a prolific source of natural products. *Tetrahedron*. 57:9347-9377.
4. Yuan, Y.V., D.E. Bone and MF. Carrington,. 2005. Antioxidant activity of dulce (*Palmariapalmata*) extract evaluated *in vitro*. *Food Chem.*, 91: 485-494.
5. Chapman V.J and D.J Chapman1980, seaweeds and their uses, 3rd edition Chapman and Hall New York. pp62-96
6. K. Renugadevi, C. ValliNachiyar, Sandeepanna, Nishantkumar, Biopotential Activity of Marine micro algae extracts, *International Journal of ChemTech Research*, Coden USA: IJCRRG, ISSN:0974-4290, Vol-6, No.12, pp-5101-5106, October 2014.
7. Silva, Basson& Moe 1996, Oliveira, Österlund&Mtolera 2005, Silva, P.C., Basson, P.W. & Moe, R.L. (1996). Catalogue of the benthic marine algae of the Indian Ocean. *University of California Publications in Botany* 79: 1-1259.
8. De Clerck, Tronchin&Schils 2005, De Clerck, O., Tronchin, E.M. &Schils, T. (2005). Red algae. Rhodophyceae. Guide to the seaweeds of KwaZulu-Natal. *ScriptaBotanicaBelgica* 33: 131-267.), Tanzania (incl. Zanzibar)
9. Silva, Basson& Moe 1996, Oliveira, Österlund & Mtolera 2005, Silva, P.C., Basson, P.W. & Moe, R.L. (1996). Catalogue of the benthic marine algae of the Indian Ocean. *University of California Publications in Botany* 79: 1-1259.
10. Silva, Basson & Moe 1996, Sahoo et al. 2001, Rao & Gupta 2015Sahoo, D., Nivedita&Debasish (2001). *Seaweeds of Indian coast*. pp. xxi + 283. New Delhi: A.P.H. Publishing. Rao, P.S.N. & Gupta, R.K. (2015). *Algae of India Volume 3. A checklist of Indian marine algae (excluding diatoms & dinoflagellates)*. pp. [i]-xviii, [1]-93, 11 pls. Salt Lake, Kolkata: Botanical Survey of India Ministry of Environment, Forests & Climate Change Government of India). Verified by: 19 July 2011 by M.D. Guiryhttp://www.algaebase.org/search/species/detail/?species_id=u8a14b0fcf292c243
11. M.D. Guiry in Guiry, M.D. &Guiry, G.M. 2016. *AlgaeBase*. World-wide electronic publication, National University of Ireland, Galway. <http://www.algaebase.org>; searched on 03 September 2016.
12. Merlin, M.J, V. Parthasarathy, R. Manavalan and S. Kumaravel 2009. Chemical Investigation of Aerial Parts of *Gmelinaasiatica* Linn by GC-MS Pharmacognosy Res, 1(3): 152-156.
13. Mohammed. A et al, Effects of aqueous extract of *GanodermaLucidum* on blood glucose levels of normoglycemic and alloxan induced diabetic wistar rats. *J. Med. Plants. Res.*2007: 1(2): 34-37.
14. Bhesh Raj S. et al Caulerpalentillifera extract ameliorates insulin resistance and regulates glucose metabolism in C57BL/KSJ-db/db mice via P13K/AKT signaling pathway in myocytes, *Journal Transl Med* 2015:13:62.

15. Murugesan S et.al Evaluation of in vitro antidiabetic activity of red sea weed *Portieriahornemanii* (Lyngbye)(Silva) and *spyridiafusiformis* (Wulfen): World journal of Pharmaceutical sciences 2016. ISSN 2321-3086.
